

3232E

**SCIENTIFIC CRITERIA DOCUMENT
FOR THE DEVELOPMENT OF
A PROVINCIAL WATER QUALITY
GUIDELINE FOR
TOLUENE**

NOVEMBER 1994



**Ministry of
Environment
and Energy**

ISBN 0-7778-3014-0

**SCIENTIFIC CRITERIA DOCUMENT FOR THE
DEVELOPMENT OF A PROVINCIAL WATER QUALITY GUIDELINE
FOR
TOLUENE**

NOVEMBER 1994



Cette publication technique
n'est disponible qu'en anglais.

Copyright: Queen's Printer for Ontario, 1994
This publication may be reproduced for non-commercial purposes
with appropriate attribution.

PIBS 3232E

**SCIENTIFIC CRITERIA DOCUMENT FOR THE
DEVELOPMENT OF A PROVINCIAL WATER QUALITY GUIDELINE
FOR
TOLUENE**

Report prepared by:

R. V. Angelow and N. Bazinet

Standards Development Branch
West Central Region
Ontario Ministry of Environment and Energy

November 1994

ACKNOWLEDGEMENTS

We gratefully acknowledge Dr. D.M. Trotter of Monenco Consultants Ltd, John Ralston, Wolfgang Scheider, Dave Rokosh, Tim Fletcher, Tom Dickie, Sum Chi Li, E. Hagmajer and other members of the MOEE Aquatic Criteria Development Committee for contributing to the development of this guideline document.

Additional review comments were received from several sources including regional and district offices, the Air Resources Branch, the Waste Management Branch, the MISA Advisory Committee, and the Advisory Committee on Environmental Standards of the Ministry of the Environment and Energy, Dr. G. Dixon (University of Waterloo), Dr. J. Meyer (University of Wyoming), and Dr. J. Giesy (Michigan State University).

The public were notified of the proposed Provincial Water Quality Guideline for toluene through the Environmental Bill of Rights Electronic Registry and given the opportunity to comment in accordance with the Environmental Bill of Rights.

PREFACE

The Ontario Ministry of Environment and Energy (MOEE) develops Provincial Water Quality Objectives or Guidelines for those substances deemed to be of environmental concern in Ontario as determined through a screening process which considered persistence, potential to bioaccumulate, acute and chronic toxicity, and potential presence in the aquatic environment. In addition, Ministry staff with a direct responsibility for managing possible effects of contaminants may request an evaluation.

Provincial Water Quality Objectives and Guidelines (PWQO/Gs) are numeric or narrative criteria intended to protect all life stages of aquatic organisms for indefinite exposures and/or they are intended to protect recreational uses of water. Objectives or guidelines do not take into account analytical detection or quantification limits, treatability or removal potential, socio-economic factors, natural background concentrations, or potential transport of contaminants among air, water and soil. They represent a desirable water quality for the protection of designated uses of surface waters in Ontario.

The process for deriving these criteria is detailed in Ontario's Water Quality Objective Development Process (1992) and is available from the ministry's Public Information Centre, 135 St. Clair Avenue, Toronto, Ontario M4V 1P5 (Tel. (416) 323-4321 or 1-800-565-4923). The toxicology literature is reviewed for all of the following areas: aquatic toxicity, bioaccumulation, mutagenicity and aesthetic considerations. The final criterion is based on the lowest effect reported for any of these. Where numeric criteria are set to protect aquatic life, the number is derived by dividing the lowest adverse effect concentration by a safety factor for Objectives or an "uncertainty factor" for Guidelines. The size of the uncertainty factor reflects the quality and quantity of data available and the potential of the material to bioaccumulate.

Policies and procedures which govern the uses of PWQO/Gs are contained in the booklet - Water Management (1984) - which deals with all aspects of Ontario's water management policy. These policies and procedures make provision for considering such factors as natural background levels, socio-economic factors, treatability, and the waste assimilative capacity of the receiving environment in applying the PWQO/Gs in site-specific situations. PWQO/Gs are used to: i) classify receiving waters for water management purposes; ii) assess contaminant discharges to the aquatic environment; and iii) derive water quality-based effluent limits which may be included in Certificates of Approval which are issued to regulate effluent discharges. Where better water quality is required to protect other beneficial uses of the environment in a given location, appropriate criteria and factors, including public health considerations, are taken into account.

EXECUTIVE SUMMARY

A Provincial Water Quality Guideline (PWQG) was developed for toluene for the protection of aquatic life. The physical-chemical properties, aquatic toxicity, bioaccumulation potential, taste and odour characteristics, and genotoxicity potential of toluene were considered in developing the guideline.

Toluene is a clear colourless liquid that occurs naturally in coal and crude oil and is found in many consumer products including gasoline, cosmetics, and cleaners. It may enter the environment from releases associated with its production, use, storage and transportation, including petroleum spills. In Ontario, the major producers of toluene are located near Sarnia. Some important dischargers of toluene into surface waters are the chemical manufacturing, petroleum refining, and pulp and paper sectors.

Natural processes can remove toluene from land, air, and water and reduce environmental exposures. As a result, ambient levels remain low and there is little tendency for toluene levels to build up in the environment over time. In Ontario, surface water concentrations are usually less than 0.001 mg/L or they are not detectable and atmospheric concentrations are generally less than 10 µg/m³. The lowest MOEE detection limit for routine analysis of toluene in water is 0.00005 mg/L.

Toluene is highly toxic to aquatic organisms when water concentrations exceed 5 mg/L during short-term exposures or reach 0.02 mg/L during long-term exposures. Reported levels of toxicity include a 96h-LC50 (median lethal concentration) of 5.46 mg/L for coho salmon fry (Moles *et al.* 1981) and a 24h-EC50 of 7 mg/L for the immobilization of water fleas (Galassi *et al.* 1988). Black *et al.* (1982) reported a 27d-LC50 of 0.02 mg/L for exposure of early life stages of rainbow trout.

The Provincial Water Quality Guideline of 0.0008 mg/L was derived by dividing the lowest adverse effect concentration (0.02 mg/L) by a final uncertainty factor of 26. Since the water quality criterion for the protection of aquatic life is more stringent than the odour, tainting and taste protection values derived for toluene, it is recommended as the PWQG.

TABLE OF CONTENTS

ACKNOWLEDGEMENTS	i
PREFACE	ii
EXECUTIVE SUMMARY	iii
TABLE OF CONTENTS	iv
 1.0 INTRODUCTION	 1
1.1 SOURCES OF TOLUENE IN THE ENVIRONMENT	2
1.2 ENVIRONMENTAL FATE AND PROPERTIES	4
 2.0 AQUATIC TOXICITY	 6
2.1 ACUTE TOXICITY	7
2.1.1 Vertebrates	7
2.1.2 Invertebrates	7
2.2 CHRONIC TOXICITY	8
2.2.1 Vertebrates	8
2.2.2 Invertebrates	9
2.2.3 Other Organisms (Plants, Protozoa, Rotatoria, and Bacteria)	9
2.3 SUMMARY OF TOXICITY DATA	9
 3.0 BIOACCUMULATION	 10
 4.0 MUTAGENICITY	 10
 5.0 ODOUR AND TASTE	 11
 6.0 DERIVATION OF WATER QUALITY GUIDELINE	 12
6.1 CALCULATION OF THE FINAL UNCERTAINTY FACTOR	12
6.2 CALCULATION OF THE GUIDELINE VALUE	15
 7.0 RESEARCH NEEDS	 16
 8.0 AMBIENT WATER QUALITY CRITERIA OF OTHER AGENCIES	 16
 9.0 REFERENCES	 17
 Table 1: Physical-Chemical Properties	 23
Table 2: Acute Toxicity Data for Toluene	24
Table 3: Chronic Toxicity Data for Toluene	31
Table 4: Uncertainty Factor Worksheet	35
Fig. 1: Toxicity Summary and Guideline Derivation Graph	36

1.0 INTRODUCTION

Toluene (C_7H_8) is a clear colourless liquid with a sweet odour. It occurs naturally in crude oil and is found in many consumer products such as gasoline, nail polish, cosmetics, rubber cement, paint brush cleaners, stain removers, fabric dyes, inks, and adhesives used for packaging of food (ATSDR 1989).

In 1982, the major use of toluene in Canada was as a solvent in consumer products, lacquers, resin solutions, lacquer thinners, and pesticides (Environment Canada 1984). Toluene was also used for manufacturing benzene and benzoic acid/phenol.

The major Ontario producers of toluene are located in Sarnia (Esso Chemical Canada and Sunchem) and Corunna (Polysar Energy and Chemical, and Shell Canada). Ontario nameplate production capacity in 1987 was 495 kilotonnes per year (Corpus Information Services Limited 1988). There are at least 20 major Ontario buyers of toluene (Corpus Information Services Limited 1988). Most buyers are located in Metropolitan Toronto with others in Oshawa, Mississauga, Cornwall, Ajax, Oakville, Brampton, Guelph, Brantford, London, and Corunna.

The effects of toluene on human health have been recently reviewed (RTECS 1993; IRIS 1992; ATSDR 1989; Health and Welfare Canada 1988). Limited health effects information is available regarding dermal exposures to aqueous solutions of toluene. The U.S. Environmental Protection Agency has given toluene a cancer rating of "D" (not classifiable as to human carcinogenicity because of a lack of human data and inadequate animal studies). The U.S. EPA has also developed an oral reference dose of 0.22 mg/kg/day (equivalent to 14 mg/day for a 70 kg person). The reference dose is an estimate of the daily exposure of the human population to a potential hazard that is likely to be without risk of deleterious effects during a lifetime.

The most likely pathway by which humans may be exposed to toluene is by breathing toluene in air (ATSDR 1989). The U.S. EPA estimated that a typical absorbed dose based on inhalation is 0.32 mg/day assuming an air exposure concentration of 34 $\mu\text{g}/\text{m}^3$. Intake from food and water may contribute smaller amounts and smoking may contribute 1 or more mg/day. Higher exposure levels might occur for individuals living near an industrial source of toluene emissions, but the significance of this can only be evaluated on a site-specific basis.

In Canada, the estimated total average daily intakes of toluene for various age groups in the population ranged from 0.0016 to 0.0216 mg/kg body weight (Government of Canada 1992). Moreover, the daily intake is considered to be well below levels that a person can be exposed to over a lifetime without deleterious effects. Hence, the Ministers of Environment Canada and Health and Welfare Canada concluded that current concentrations of toluene in the environment do not constitute a danger in Canada to human life or health.

The purpose of this document is to develop a Provincial Water Quality Guideline for toluene for the protection of aquatic life. Toluene was identified as a priority for guideline development because it was deemed to be a substance of environmental concern and was present in industrial point source discharges to Ontario surface waters. In addition, a guideline for the protection of aquatic life was developed because toluene is a common pollutant released into water through chemical spills.

1.1 SOURCES OF TOLUENE IN THE ENVIRONMENT

Toluene may enter the environment from discharges or spills (including petroleum spills) associated with its use, production, storage and transportation (Fishbein 1985). Atmospheric emissions are expected from industries by way of process and fugitive emissions and evaporation from wastewater streams. Also, it is anticipated that process residues and sludges containing these chemicals may enter landfills.

Toluene has been detected in the effluents of various industries monitored in Ontario under the MISA (Municipal Industrial Strategy for Abatement) Program. These industries were associated with the Organic Chemical Manufacturing (OCM), Petroleum Refining, Inorganic Chemicals, Pulp and Paper, Iron and Steel, Metal Casting, Industrial Minerals, and Metal Mining sectors.

In the OCM sector, toluene was detected in effluents indicating point source inputs to the St. Clair and St. Lawrence rivers and in the vicinity of Elmira and Thorold. Average concentrations ranged from 0.0004 to 0.050 mg/L among the important dischargers to the environment located on the St. Clair River (MOEE unpublished data, 1989-1991). The total sector loading of toluene is approximately 6 kg/day.

In the Petroleum sector, toluene was detected in 16 percent of the samples for all 7 refineries located in Ontario. The average concentration was 0.0004 mg/L indicating potential inputs to the St. Clair River near Sarnia, Lake Erie near Nanticoke, and Lake Ontario near Oakville (MOE 1990).

Average concentrations of toluene detected in discharges for the Inorganic Chemicals sector ranged up to 0.003 mg/L indicating inputs to the St. Clair River near Sarnia, the Detroit River at Amherstburg, the Welland River, Lake Gibson (Thorold), and the St. Lawrence River at Maitland and Cornwall (MOEE unpublished data, 1990-1991).

Average concentrations of toluene detected in Pulp and Paper sector effluents ranged from 0.00013 to 0.023 mg/L in 1990 (Environment Ontario 1991). In 1990, there were 27 pulp and paper mills located in Ontario in areas such as Fort Francis, Dryden, Thunder Bay, Marathon, Espanola, Iroquois Falls, St. Catharines, Trenton, Ottawa, and Cornwall.

The average concentration of toluene detected in Iron and Steel sector effluents ranged from 0.0004 to 0.0034 mg/L indicating inputs to Hamilton harbour and the Ottawa river during 1990-91 (Hamdy 1991). Also, toluene was detected in the Metal Casting, Industrial Minerals,

and Metal Mining sectors at levels generally less than 0.001 mg/L (MOEE unpublished data, 1990-91).

1.2 ENVIRONMENTAL FATE AND PROPERTIES

The physical and chemical properties of toluene are shown in Table 1. Toluene is moderately soluble in water. The high Henry's Law Constant and vapour pressure indicate toluene is highly volatile.

In the aquatic environment, toluene will be removed by both volatilization and biodegradation (Howard 1990; Callahan et al. 1979). The predominant removal process will depend on water temperature, mixing conditions, and the existence of acclimated microorganisms at the site. The overall half-life of toluene in surface water may range from days to several weeks (Howard 1990). The half-life for volatilization of toluene from a water column one metre deep has been reported to range between 30 minutes and 5 hours (Mackay and Wolkoff 1973; Mackay and Leinonen 1975; Thomas 1982).

Toluene is readily degradable in a variety of standard biodegradability tests (reviewed by Howard 1990). For example, after seven days at 23°C, a concentration of less than 5 mg/L of toluene remained in an industrial microbial test system initially dosed with 50 mg/L (Davis et al. 1981).

Toluene will not significantly hydrolyze, photolyze, adsorb to sediment, or bioconcentrate in aquatic organisms (Howard 1990). The potential for toluene to bioaccumulate is discussed further in section 3.0.

When released onto soil, toluene will evaporate into the atmosphere and may leach into groundwater, especially in soil with low organic carbon content (Howard 1990). However, toluene may be relatively persistent in groundwater where volatilization is not a viable process. Also, biodegradation will occur although the extent will depend on the microbes

present, the concentration of toluene, the presence of other compounds, and the amount of oxygen present.

Once in the atmosphere, toluene will be transported until it is removed by physical (i.e., partitioning into clouds or rainwater) or chemical processes (ATSDR 1989). Toluene will principally be removed by chemical transformations caused by the sun's energy (photooxidation) with a half-life of less than 2 days (ATSDR 1989; Howard 1990).

Toluene has been detected throughout North America in effluents from municipalities and industries, industrialized river basins, groundwater, sediments, soil and air (ATSDR 1989; Howard 1990; Fishbein 1985). Despite its widespread presence, physical, chemical and biological processes can remove toluene in all media and reduce environmental exposures. Hence, ambient levels remain low because of photooxidation in air and high volatility from water and soil.

In 1986, a survey was undertaken to investigate the potential impact of point source discharges to the St. Clair River (Environment Ontario 1991). Water samples were collected from 43 stations located throughout the river in May, July, and October. Toluene was not detected in 284 samples collected at the detection limit of 0.001 mg/L.

As part of the Ontario Drinking Water Surveillance Program which includes raw water samples collected from surface and ground waters from urbanized areas located throughout the Great Lakes basin, toluene was usually not detectable (MOEE unpublished data, 1992). Toluene was detected at a few sites at levels up to 0.00025 mg/L. The MOEE routine analytical detection limit for toluene in water is 0.00005 mg/L. The Canadian Drinking Water Guideline for public water supplies is 0.024 mg/L (aesthetic objective).

In the United States, toluene was detected in soil samples collected at hazardous waste sites studied by the Environmental Protection Agency at levels averaging 0.077 mg/kg (ATSDR

1989). In a survey of 7 landfill sites tested in Ontario, toluene was detected in leachate at levels ranging from 0.001 to 0.6 mg/L (MOEE unpublished data, 1988).

As part of the MOEE Volatile Organic Compounds Monitoring Network, ambient air concentrations of toluene for six cities (Thunder Bay, Sault Ste. Marie, Windsor, Toronto, and Hamilton) ranged from not-detectable (less than $0.1 \mu\text{g}/\text{m}^3$) to $52 \mu\text{g}/\text{m}^3$ (Environment Ontario 1992). This upper level was found in Windsor, although the average concentration here was $12 \mu\text{g}/\text{m}^3$. In general, measured concentrations were less than $10 \mu\text{g}/\text{m}^3$. The Ontario ambient air quality criterion or acceptable level is $2000 \mu\text{g}/\text{m}^3$.

In summary, toluene is rapidly degraded by photochemical oxidation in the atmosphere. Toluene in water or soil will volatilize to air and that which remains is subject to microbial degradation. As a result, there is little tendency for toluene levels to build up in the environment over time.

2.0 AQUATIC TOXICITY

Criteria used for classifying available toxicity data as either primary or secondary information are described in "Ontario's Water Quality Objective Development Process" (Environment Ontario 1992). In general, primary toxicity studies involve acceptable test procedures, conditions, and controls, measured toxicant concentrations, and flow through or renewal exposure conditions. Secondary toxicity studies usually involve unmeasured toxicant concentrations, static bioassay conditions, and unsatisfactory reporting of experimental data. Generally, acute toxicity studies involve test durations of 96 hours or less for vertebrates or 48 hours or less for invertebrates. Chronic toxicity data studies include complete life cycle tests and partial life cycle tests involving early life stages.

2.1 ACUTE TOXICITY

2.1.1 Vertebrates

Primary acute toxicity data for animals exposed to toluene were available for early life stages of fathead minnow (Pimephales promelas), goldfish (Carassius auratus), guppy (Lebistes reticulatus), rainbow trout (Oncorhynchus mykiss) and coho salmon (Oncorhynchus kisutch) fry (Table 2). Primary 96h-LC50 values ranged from 5.46 mg/L for coho salmon fry (Moles et al. 1981) to 72 mg/L for fathead minnow embryos (Devlin et al. 1982). The lowest reported 96h-LC50 values for rainbow trout and fathead minnow fry were 5.8 and 18 mg/L, respectively (Galassi et al. 1988; Devlin et al. 1982).

Secondary acute toxicity data were available for fathead minnow, goldfish, rainbow trout, bluegill sunfish (Lepomis macrochirus), guppy, channel catfish (Ictalurus punctatus), golden orfe (Leuciscus idus), and mosquito fish (Gambusia affinis) (Table 2). 96h-LC50 values ranged from 12.6 mg/L for fathead minnow (Liu et al. 1983) to 1180 mg/L for mosquito fish (Wallen et al. 1957). It is important to note that the 24, 48 and 96h-LC50 values reported by Wallen et al. (1957) exceed the solubility of toluene in pure water.

2.1.2 Invertebrates

In a study considered as primary data, Galassi et al. (1988) reported a 24h-EC50 of 7 mg/L for the immobilization of water fleas (Daphnia magna). Also, secondary acute toxicity data were available for Daphnia magna. LC50 or EC50 values for Daphnia magna ranged from 11.5 mg/L (Bobra et al. 1983) to 310 mg/L (LeBlanc 1980).

2.2 CHRONIC TOXICITY

2.2.1 Vertebrates

Primary chronic toxicity data were available for coho salmon, rainbow trout, goldfish, fathead minnow, salamander (Ambystoma gracile), and leopard frog (Rana pipiens) (Table 3). Moles et al. (1981) reported effect concentrations of 2.76 and 5.00 mg/L for significantly reduced growth of coho salmon fry after 40 days of exposure. LC50 values, derived from long term exposures, ranged from 0.02 mg/L for rainbow trout (Black et al. 1982) to 44.1 mg/L for fathead minnow (Hall et al. 1984,1989). Brenniman et al. (1976) reported a 30d-LC50 of 14.58 mg/L for goldfish. Hall et al. (1984, 1989) reported an 8d-LC50 of 44.1 mg/L for fathead minnow. In addition, Black et al. (1982) reported median lethal concentrations of 0.85 and 1.09 mg/L for early life stages of salamander and median lethal concentrations of 0.39 and 0.51 mg/L for early life stages of frogs.

In studies considered secondary data, Stoss and Haines (1979) reported a 96h-LC50 of 54 mg/L for medaka embryos (Oryzias latipes) and Könemann (1981) reported a 14d-LC50 of 68.3 mg/L for guppy (Poecilia reticulata).

In a study considered as ancillary information, Devlin et al. (1985) exposed fathead minnow embryos (1-2 hours old) to 30 to 45 mg/L of toluene for up to 124 hours. The chorion of toluene-treated embryos were found to be cloudy, fragile and covered with a white gelatinous material. Embryo-toxic effects noted included a curved embryonic axis, abnormal heart and circulatory system development, hydration and swelling of the pericardial coelom, haemorrhaging, microphthalmia, enlarged yolk sac, and an overall stunted appearance and retarded development. Virtually all tissues of toluene exposed embryos were abnormal.

2.2.2 Invertebrates

No chronic toxicity studies for exposed invertebrates were available. However, Hermens et al. (1984) calculated a 16d-LC50 of 3.75 mg/L based on quantitative structure-activity relationships (QSAR).

2.2.3 Other Organisms (Plants, Protozoa, Rotatoria, and Bacteria)

As defined by Environment Ontario (1992), secondary chronic toxicity data were available for algae, rotatoria, and protozoa (Table 3). The lowest reported adverse effect concentration was a 72h-EC50 of 12.5 mg/L for algae, Selenastrum capricornutum (Galassi et al. 1988). Erben (1978) found an EC40 of approximately 173 mg/L for rotifers (Dicranophorus forcipatus). In addition, Schultz et al. (1978) found a 24h-LC100 of 550 mg/L for protozoa (Tetrahymena pyriformis).

2.3 SUMMARY OF TOXICITY DATA

The ranges of toxicity exhibited by aquatic organisms exposed to toluene are summarized in Figure 1. Acute toxicity data were available for both cold water and warm water fish species. Primary 96h-LC50 values ranged from 5.46 mg/L for coho salmon fry (Moles et al. 1981) to 72 mg/L for fathead minnow embryos (Devlin et al. 1982).

Limited primary acute toxicity information was available for invertebrates. Galassi et al. (1988) reported a 24h-EC50 of 7 mg/L (immobilization) for water fleas.

Primary chronic toxicity data were available for cold and warm water fish species and early life stages of frogs and salamanders. LC50 values, derived from long term exposures, ranged from 0.02 mg/L for early life stages of rainbow trout (Black et al. 1982) to 44.1 mg/L for fathead minnows (Hall et al. 1984, 1989).

No chronic toxicity data were found for macro-invertebrates exposed to toluene. The lowest reported adverse effect concentrations for algae (Selenastrum capricornutum), rotifers (Dicranophorus forcipatus), and protozoa (Tetrahymena pyriformis) were 12.5 mg/L (72h-EC50), approximately 173 mg/L (6d-EC40), and 550 mg/L (24h-LC100), respectively (Galassi et al. 1988; Erben 1978; Schultz et al. 1978).

3.0 BIOACCUMULATION

The log Kow for toluene is 2.7 which is characteristic of a compound with a low potential for bioaccumulation (Hawker and Connel 1988; Lyman et al. 1982). Veith et al. (1980) calculated a bioconcentration factor (BCF) of between 15 and 70. The alga, Chlorella fusca, concentrated toluene by a factor of 380 (Geyer et al. 1984). In conclusion, toluene is not expected to undergo significant bioaccumulation in aquatic organisms.

4.0 MUTAGENICITY

The genotoxic effects of toluene have been recently reviewed (IRIS 1992; GENETOX 1992; ATSDR 1989; Fishbein 1985). In summary, genotoxicity studies have provided negative results in a variety of assays using bacteria (Salmonella and Escherichia coli strains) in the presence and absence of metabolic activation or yeast (Saccharomyces cerevisiae). Administration of toluene to mice did not induce dominant lethal mutations in sperm cells. There was no evidence of chromosomal aberrations in blood lymphocytes of workers exposed to toluene only and there was no evidence of chromosomal aberrations or sister chromatid exchanges in cultured human lymphocytes exposed to toluene in vitro. However, two isolated studies reported that toluene was effective in causing chromosomal damage in bone marrow cells of rats (Lyapkalo 1973; Dobrokhotov 1972).

A review of the scientific literature indicated there was no mutagenicity or genotoxicity information available for aquatic plants and animals exposed to toluene. As a result, there is

insufficient information available to develop a numerical criterion for the protection of aquatic life.

Although the majority of the data suggest that toluene is not genotoxic, the two studies that did show positive results suggest that toluene may have the potential for causing genotoxicity in aquatic organisms.

5.0 ODOUR AND TASTE

Zoeteman et al. (1971) reported a threshold odour concentration (T.O.C.) of 1 mg/L for toluene in water at 15 °C. Toluene in water was reported to taint or impair the taste of yellow perch (Perca flavatilis) at concentrations ranging from 0.25 to 50 mg/L (as reviewed by Persson 1984).

Alexander et al. (1982) measured aqueous taste and odour thresholds for toluene in odour-free water. The average taste threshold was 0.14 mg/L at 40 °C. The average odour threshold was 0.024 mg/L at 60 °C. However, this odour threshold value is not considered representative of ambient conditions because aquatic temperatures are usually much lower. Also, this study showed that odour threshold values for several volatile substances were consistently higher at lower temperatures (i.e., 20 °C).

6.0 DERIVATION OF WATER QUALITY GUIDELINE

Since the toxicological database for toluene was limited, a Provincial Water Quality Objective could not be developed. Therefore, following standard procedures as outlined in Environment Ontario (1992), the process reverted to the derivation of a Provincial Water Quality Guideline for the protection of aquatic life.

Where guidelines are set to protect aquatic life, the number is derived by dividing the lowest adverse effect concentration by an "uncertainty factor". The size of the uncertainty factor reflects the quality and quantity of data available and the potential of the material to bioaccumulate.

The Federal-Provincial Working Group on recreational water quality has not recommended limits for chemicals in recreational water for human exposure because of the lack of sufficient scientific information (Health and Welfare Canada 1992). Therefore, a recreational use water quality guideline for the protection of human health is not recommended at this time.

Humans exposed to aqueous solutions of toluene may absorb toluene through the skin (ATSDR 1989). Brown et al. (1984) estimated that concentrations of toluene at 0.005 to 0.5 mg/L would result in daily doses of 0.0002 to 0.02 mg/kg body weight (equivalent to 0.0014 to 1.4 mg per day for a 70 kg adult) while swimming for 15 minutes and consuming contaminated water. Since toluene in Ontario surface waters is present at much lower concentrations (i.e., less than 0.001 mg/L) or is not detectable, this suggests that exposure through skin contact with water is likely insignificant.

6.1 CALCULATION OF THE FINAL UNCERTAINTY FACTOR

The choice of a baseline uncertainty factor depends upon the octanol-water partition coefficient (Kow) of the substance (which is a useful indicator of bioaccumulation potential). Chiou and Schmedding (1982) reported a measured log Kow of 2.7 for toluene and Mabey et

al. (1982) reported an estimated log Kow of 2.8. Therefore, a baseline uncertainty factor of 1000 was selected, since the measured log Kow for toluene was less than 4 (Environment Ontario 1992).

The final uncertainty factor was calculated based on the following toxicity information (Table 4):

- A. The following studies were used in the chronic toxicity data category:
1. The 30-day LC50 of 14.58 mg/L for goldfish (C. auratus) was considered primary chronic toxicity information (Brenniman et al. 1976). The fish were exposed under continuous flow conditions with measured toxicant concentrations.
 2. The 27-day LC50 of 0.02 mg/L for rainbow trout (Oncorhynchus mykiss) embryos was considered as primary toxicity information (Black et al. 1982). Toxicant concentrations were measured throughout this test which involved continuous flow exposure conditions and covered test chambers.
 3. The lowest observed effect concentration of 2.76 mg/L for significantly reduced growth of Coho salmon (Oncorhynchus kisutch) was considered as primary toxicity information (Moles et al. 1981). This study involved a flow-through system with measured toxicant concentrations.
 4. The estimated 16-day LC50 of 3.75 mg/L for cladoceran (D. magna), based on quantitative structure-activity relationships, was considered tertiary chronic toxicity information (Hermens et al. 1984).
 5. The 6d-EC40 (cell culture growth) of approximately 173 mg/L for the rotifer (Dicranophorus forcipatus) was considered secondary toxicity information and used in the invertebrate category as defined by Environment Ontario (1992). This study

involved unmeasured toxicant concentrations and static exposure conditions (Erben 1978).

6. The 72h-EC50 (growth) of 12.5 mg/L for algae (Selenastrum capricornutum) was considered secondary information (Galassi et al. 1988). This study involved static conditions and measured toxicant concentrations.

B. The following studies were used in the acute toxicity data category:

1. The 96h-LC50 of 18 mg/L for fathead minnow (Pimephales promelas) was considered primary acute toxicity information (Devlin et al. 1982). This study involved flow through conditions and measured toxicant concentrations.
2. The 96h-LC50 of 5.8 mg/L for rainbow trout was considered primary acute toxicity information (Galassi et al. 1988). This test involved renewal exposure conditions in sealed containers and measured toxicant concentrations.
3. The 96h-LC50 of 5.46 mg/L for coho salmon fry (Oncorhynchus kisutch) was considered primary toxicity information (Moles et al. 1981). This test involved flow through conditions and measured toxicant concentrations.
4. The 24h-EC50 (immobilization) of 7.0 mg/L for 24 hour old cladoceran (D. magna) was considered primary toxicity information (Galassi et al. 1988). This test involved static exposure conditions in closed bottles and measured toxicant concentrations.

Based on the above data and applying the appropriate calibration factors (Table 4), a value of 26 was derived as the final uncertainty factor.

6.2 CALCULATION OF THE GUIDELINE VALUE

The following guideline was set as a single value independent of other water quality parameters such as temperature. Since toluene is highly volatile, only primary studies were considered in selecting the lowest adverse effect concentration or critical value.

The lowest adverse effect concentration considered valid for developing a PWQG was a 27-day LC50 of 0.02 mg/L reported for rainbow trout in an embryo-larval test (Black *et al.* 1982). This value divided by the final uncertainty factor of 26 produced a preliminary Provincial Water Quality Guideline of 0.0008 mg/L (rounded to one significant figure) (Table 4 and Fig. 1).

For toluene in water, the lowest valid threshold odour concentration was 1 mg/L (Zoeteman *et al.* 1971). This value multiplied by a safety factor of 0.5 produced an odour protection value of 0.5 mg/L. Similarly, a tainting protection value of 0.13 mg/L and a taste protection value of 0.070 mg/L were derived by applying a safety factor of 0.5 to water concentrations found to impair the taste of yellow perch (0.25 mg/L) and the taste of water (0.14 mg/L), respectively (Section 5.0).

There was no information available to develop a numerical mutagenicity criterion for the protection of aquatic life, since no acceptable genotoxicity information for fish, invertebrates, or plants was found. Most of the available data suggest toluene is not genotoxic, although chromosomal damage by this compound was reported (Section 4.0). While the majority of evidence supports the classification of toluene as non-mutagenic, the possibility of its genotoxic hazard in aquatic organisms cannot be excluded.

Since the preliminary guideline value of 0.0008 mg/L based on toxicity is below the odour, tainting and taste protection values, the recommended Provincial Water Quality Guideline for the protection of aquatic life is 0.0008 mg/L. The MOEE routine analytical detection limit for toluene in water is 0.00005 mg/L ("clean" water analysis).

7.0 RESEARCH NEEDS

Additional toxicity studies involving freshwater macro-invertebrates are needed to fulfil the minimum data requirements for developing a PWQO.

8.0 AMBIENT WATER QUALITY CRITERIA OF OTHER AGENCIES

The CCREM (1987) recommended a Canadian water quality guideline of 0.3 mg/L for the protection of aquatic life.

The U.S. EPA has not developed a water quality criterion for the protection of freshwater biota, although they report 17.5 mg/L is the lowest effect concentration (LEC) for acute toxicity (IRIS 1992).

9.0 REFERENCES

- Alexander, H.C., W.M. McCarty, E.A. Bartlett and A.N. Syverud. 1982. Aqueous odor and taste threshold values of industrial chemicals. J. Am. Water Works Assoc. 74: 595-599.
- ATSDR. 1989. Agency for Toxic Substances and Disease Registry. Toxicological Profile for toluene. U.S. Public Health Service. December, 1988. 115 pp.
- Black, J.A., W.J. Birge, W.E. McDonnell, A.G. Westerman, B.A. Ramey and D.M. Bruser. 1982. The Aquatic Toxicity of Organic Compounds to Embryo-Larval Stages of Fish and Amphibians. University of Kentucky Water Resources Research Institute. Research Report No. 133.
- Bobra, A.M., W.Y. Shiu and D. Mackay. 1983. A predictive correlation for the acute toxicity of hydrocarbons and chlorinated hydrocarbons to the water flea, Daphnia magna. Chemosphere 12: 1121-1129.
- Brenniman, G., R. Hartung and W.J. Weber, Jr. 1976. A continuous flow bioassay method to evaluate the effects of outboard motor exhausts and selected aromatic toxicants on fish. Water Research 10: 165-169.
- Bridie, A.L., C.J.M. Wolff and M. Winter. 1979. The acute toxicity of some petrochemicals to goldfish. Water Research 13: 623-626.
- Bringmann, G. and R. Kuhn. 1980. Comparison of the toxicity thresholds of water pollutants to bacteria, algae and protozoa in the cell multiplication inhibition test. Water Res. 14: 231-241.
- Brown, H.S., D.R. Bishop and C.S. Rowan. 1984. The role of skin absorption as a route of exposure for volatile organic compounds (VOCs) in drinking water. American Journal of Public Health 74(5): 479-484.
- Buccafusco, R.J., S.J. Ells and G.A. LeBlanc. 1981. Acute toxicity of priority pollutants to Bluegill (Lepomis macrochirus). Bull. Environ. Contam. Toxicol. 26: 446-452.
- Callahan, M.A., M.W. Slimak, N.W. Gabel, I.P. May, C.F. Fowler, J.R. Freed, P. Jennings, R.L. Durfee, F.C. Whitmore, B. Maestri, W.R. Mabey, B.R. Holt and C. Gould. 1979. Water-Related Environmental Fate of 129 Priority Pollutants. Volume II: Halogenated Aliphatic Hydrocarbons, Halogenated Ethers, Monocyclic Aromatics, Phthalate Esters, Polycyclic Aromatic Hydrocarbons, Nitrosamines, and Miscellaneous Compounds. EPA 440/4-79-029b.

- CCREM. 1987. Canadian Water Quality Guidelines. Task Force on Water Quality Guidelines of the Canadian Council of Resource and Environment Ministers, March, 1987.
- Chiou, C.T. and D.W. Schmedding. 1982. Partitioning of organic compounds in octanol-water systems. Environ. Sci. Technol. 16: 4-10.
- Corpus Information Services Limited. 1988. Chemical Production Profiles - Toluene. Chemical Product Profiles, Don Mills, Ontario. (November, 1987).
- Davis, E.M., H.E. Murray, J.G. Liehr and E.L. Powers. 1981. Basic microbial degradation rates and chemical byproducts of selected organic compounds. Water Research 15: 1125-1127.
- Dobrokhotov, V.B. 1972. The mutagenic influence of benzene and toluene under experimental conditions. Gig. Sanit. 37:36-39. (Russian).
- Devlin, E.W., J.D. Brammer and R.L. Puyear. 1982. Acute toxicity of toluene to three age groups of fathead minnows (Pimephales promelas). Bulletin of Environmental Contamination and Toxicology 29: 12-17.
- Devlin, E.W., J.D. Brammer and R.L. Puyear. 1985. Effect of toluene on fathead minnow (Pimephales promelas Rafinesque) development. Arch. Environ. Contam. Toxicol. 14: 595-603.
- Environment Canada. 1984. Toluene - Environmental and Technical Information for Problem Spills. Technical Services Branch. Ottawa, Ontario. 104 pp.
- Environment Ontario. 1984. Water Management. Goals, Policies, and Implementation Procedures of the Ministry of the Environment. Toronto, Ontario. 70 pp.
- Environment Ontario. 1991. St. Clair River MISA Pilot Site Investigation. Volume II. Part II. Detailed Technical Findings. Prepared by the St. Clair River MISA Pilot Site Team. Environment Ontario. Water Resources Branch. Toronto, Ontario.
- Environment Ontario. 1991. Preliminary Report on the First Six Month of Process Effluent Monitoring in the MISA Pulp and Paper Sector (January 1, 1990 to June 30, 1990). Environment Ontario. Water Resources Branch. Toronto, Ontario. 119 pp.
- Environment Ontario. 1992. Ontario's Water Quality Objective Development Process. Ontario Ministry of the Environment. Water Resources Branch. 56 pp.

- Environment Ontario. 1992. Volatile Organic Compounds Monitoring Network. Ambient Air Concentration Data Listing 1990. Ontario Ministry of the Environment. Air Resources Branch. 24 pp.
- Erben, R. 1978. Effects of some petrochemical products on the survival of Dicranophorus forcipatus O.F. MULLER (Rotatoria) under laboratory conditions. Verh. Internat. Verein. Limnol. 20:1988-1991.
- Fishbein, L. 1985. An overview of environmental and toxicological aspects of aromatic hydrocarbons. II. Toluene. Sci. Tot. Environ. 42: 267-288.
- Galassi, S., M. Mingazzini, L. Viagno, D. Cesareo and M.L. Tosato. 1988. Approaches to modelling toxic responses of aquatic organisms to aromatic hydrocarbons. Ecotoxicol. Environ. Saf. 16(2): 158-169.
- Geiger, D.L., S.H. Poirier, L.T. Brooke and D.J. Call (editors). 1986. Acute Toxicities of Organic Chemicals to Fathead Minnows (Pimephales promelas). Volume III. Centre for Lake Superior Environmental Studies, University of Wisconsin - Superior.
- Geiger, D.L., L.T. Brooke, and D.J. Call. 1990. Acute Toxicities of Organic Chemicals to Fathead Minnow (Pimephales promelas). Volume V. Center for Lake Superior Environmental Studies. University of Wisconsin-Superior. Superior, WI.
- GENETOX. 1992. United States Environmental Protection Agency Genetox Program. Provided by NLM Bethesda, MD. (online database).
- Geyer, H., G. Politzki and D. Freitag. 1984. Prediction of ecotoxicological behaviour of chemicals: relationship between n-octanol/water partition coefficient and bioaccumulation of organic chemicals by alga Chlorella. Chemosphere 13(2): 269-284.
- Government of Canada. 1992. Canadian Environmental Protection Act, Priority Substances List Assessment Report No. 4: Toluene. Health and Welfare Canada and Environment Canada. Ottawa. 26 pp.
- Hall, L.H., L.B. Kier and G. Phipps. 1984. Structure-activity relationship studies on the toxicities of benzene derivatives. I. An additivity model. Environ. Toxicol. Chem. 3:355-365.
- Hall, L.H., E.L. Maynard and L.B. Kier. 1989. QSAR investigation of benzene toxicity to fathead minnow using molecular connectivity. Environ. Toxicol. Chem. 8(9):783-789.
- Hamdy, Y. 1991. Status Report on the Effluent Monitoring Data for the Iron and Steel Sector for the Period from November 1, 1989 to October 31, 1990. Ontario Ministry of the Environment, Water Resources Branch. 219 pp.

- Hawker, D.W. and D.W. Connell. 1988. Influence of partition coefficient of lipophilic compounds on bioconcentration kinetics with fish. *Water Res.* 22: 701-702.
- Health and Welfare Canada. 1988. Guidelines for Canadian Drinking Water Quality. Part II. Supporting Documentation. Toluene, Ethylbenzene and the Xylenes. Ottawa, Ontario, Canada.
- Health and Welfare Canada. 1992. Guidelines for Canadian Recreational Water Quality. Ottawa, Ontario, Canada. 101 pp.
- Hermens, J., H. Canton, P. Janssen and R. DeJong. 1984. Quantitative structure-activity relationships and toxicity studies of mixtures of chemicals with anaesthetic potency: acute lethal and sublethal toxicity to Daphnia magna. *Aquatic Toxicology* 5: 143-154.
- Howard, P.H. 1990. Handbook of Environmental Fate and Exposure Data for Organic Chemicals. Volume II. Solvents. Lewis Publishers. Chelsea, MI. 546 p.
- Hutchinson, T.C., J.A. Hellebust, D. Tam, D. MacKay, R.A. Mascarenhas and W.Y. Shiu. 1980. The correlation of the toxicity to algae of hydrocarbons and halogenated hydrocarbons with their physical-chemical properties. In: Hydrocarbons and Halogenated Hydrocarbons in the Aquatic Environment. B.K. Afgan and D. MacKay (eds.) Plenum Press, N.Y.
- IRIS. 1992. Integrative Risk Information System. United States Environmental Protection Agency. Provided by NLM Bethesda, MD (online database).
- Johnson, W.W. and M.T. Finley. 1980. Handbook of Acute Toxicity of Chemicals to Fish and Aquatic Invertebrates. U.S. Department of the Interior, Fish and Wildlife Service, Resource Publication 137.
- Juhnke, V. I. and D. Lüdemann. 1978. Ergebnisse der untersuchung von 200 chemischen verbindungen auf akute fischtoxizität mit dem goldorfeentest. *T. Wasser Abwasser Forsch.* 11(5): 161-164.
- Kauss, P.B. and T.C. Hutchinson. 1975. The effects of water-soluble petroleum components on the growth of Chlorella vulgaris Beijerinck. *Environmental Pollution* 9: 157-174.
- Könemann, H. 1981. Quantitative structure-activity relationships in fish toxicity studies. Part 1: Relationship for 50 industrial pollutants. *Toxicology* 19: 209-221.
- LeBlanc, G.A. 1980. Acute toxicity of priority pollutants to water flea (Daphnia magna). *Bull. Environ. Contam. Toxicol.* 24: 684-691.

- Liu, D.H.W., H.C. Bailey and J.G. Pearson. 1983. Toxicity of complex munitions wastewater to aquatic organisms. In: Aquatic Toxicology and Hazard Assessment: Sixth Symposium. ASTM STP 802. W.E. Bishop et al. (eds.). American Society for Testing and Materials, Philadelphia, PA. p. 135-150.
- Lyapkalo, A.A. 1973. Genetic activity of benzene and toluene. *Gig. Tr. Prof. Zabol.* 17(3):24-28. (Russian).
- Lyman, W.J., W.F. Reehl and D.H. Rosenblatt. 1982. Handbook of Chemical Property Estimation Methods, Environmental Behavior of Organic Compounds. McGraw-Hill Book Co., Toronto.
- Mabey, W.R., J.H. Smith, R.T. Podell, H.L. Johnson, T. Mill, T.W. Chou, I.W. Partridge and D. Vandenberg. 1982. Aquatic Fate Process Data for Organic Priority Pollutants. EPA 440/4-81-014.
- Mackay, D. and A.W. Wolkoff. 1973. Rate of evaporation of low-solubility contaminants from water bodies to atmosphere. *Environmental Science and Technology* 7: 611-614.
- Mackay, D. and P.J. Leinonen. 1975. Rate of evaporation of low-solubility contaminants from water bodies to atmosphere. *Environmental Science Technology* 9: 1178-1180.
- Mann, H. 1953. Ueber Geschmacksbeeinflussungen bei Fishen. *Fischwirt.* 3: 330-334. (as cited in Persson 1984).
- Mayes, M.A., H.C. Alexander and D.C. Dill. 1983. A study to assess the influence of age on the response of fathead minnows in static acute toxicity tests. *Bulletin of Environmental Contamination and Toxicology* 31: 139-147.
- MOE. 1990. MISA Second Report on the Monitoring Data for the Petroleum Refining Sector. Ontario Ministry of the Environment. Water Resources Branch.
- Moles, A., S. Bates, S. D. Rice, and S. Corn. 1981. Reduced growth of Coho salmon fry exposed to two petroleum components, toluene and naphthalene, in freshwater. *Transaction of the American Fisheries Society* 110:430-436.
- Moles, A. 1980. Sensitivity of parasitized coho salmon fry to crude oil, toluene, and naphthalene. *Transactions of the American Fisheries Society* 109: 293-297.
- Persson, P-E. 1984. Uptake and release of environmentally occurring odorous compounds by fish. A review. *Water Res.* 18: 1263-1271.
- Pickering, Q.H. and C. Henderson. 1966. Acute toxicity of some important petrochemicals to fish. *J. Water Pollut. Control Fed.* 38: 1419-1429.

- RTECs. 1993. Registry of Toxic Effects of Chemical Substances. National Institute for Occupational Safety and Health. Provided by: Canadian Centre for Occupational Health and Safety (CCINFODisc.).
- Schultz, T.W., L.M. Kyte and J.N. Dumont. 1978. Structure-toxicity correlations of organic contaminants in aqueous coal-conversion effluents. Archives of Environmental Contamination and Toxicology 7: 457-463.
- Sittig, M. 1981. Handbook of Toxic and Hazardous Chemicals and Carcinogens. 2nd F.D. Noyes Publications. Park Ridge, New Jersey, U.S.A. 950 p.
- Stoss, F.W. and T.A. Haines. 1979. The effects of toluene on embryos and fry of the Japanese medaka *Oryzias latipes* with a proposal for rapid determination of maximum acceptable toxicant concentration. Environ. Pollut. 20(2):139-148.
- Thomas, R.G. 1982. Volatilization from Water. In: Handbook of Chemical Property Estimation Methods, Environmental Behaviour of Organic Compounds. W.J. Lyman et al. (eds.). McGraw-Hill Book Company, Toronto. Chapter 15, pp. 15-1 to 15-34.
- Veith, G.D., K.J. Macek, S.R. Petrocelli and J. Carroll. 1980. An evaluation of using partition coefficients and water solubility to estimate bioconcentration factors for organic chemicals in fish. J. Fish. Res. Board Can. 36:1040-1048.
- Verschueren, K. 1983. Handbook of Environmental Data on Organic Chemicals. Van Nostrand Reinhold Company, Toronto. Second Edition. 1310 pp.
- Wallen, I.E., W.C. Greer, and R. Lasater. 1957. Toxicity to Gambusia affinis of certain pure chemicals in turbid waters. Sewage and Industrial Wastes 29(6): 695-711.
- Zoeteman, B.C.J., A.J.A. Kraayeveld and G.J. Piet. 1971. Oil pollution and drinking water odour. H₂O 4(16): 367-371.

TABLE 1: PHYSICAL-CHEMICAL PROPERTIES

CHEMICAL: Toluene	CHEMICAL FORMULA: C₇H₈	CAS No: 108-88-3
-----------------------------	--	----------------------------

PROPERTIES

MOLECULAR WEIGHT (MW):	92.1 g/mol	Verschueren (1983)
MELTING POINT:	- 95.1 °C	Verschueren (1983)
BOILING POINT:	110.8°C	Verschueren (1983)
PHYSICAL STATE AT STANDARD TEMPERATURE AND PRESSURE:	liquid	
DENSITY (D):	0.867 g/cm ³ (20°/4°C)	Verschueren (1983)
MOLAR VOLUME (MW/D):	106.23 cm ³ /mol (calc.)	
VAPOUR PRESSURE (Ps):	2933 Pa at 20°C	Verschueren (1983)
WATER SOLUBILITY (Cs):	515 mg/L at 20°C	Verschueren (1983)
HENRY'S LAW CONST.(Ps/Cs):	0.006 4 atm - m ³ /mol	Howard (1990)

PERSISTENCE

SURFACE WATER HALF LIFE:	days to weeks	Howard (1990)
AQUATIC FATE:	volatilization and biodegradation	Howard (1990)

OCTANOL-WATER PARTITION COEFFICIENT (Kow)

RANGE OF AVAILABLE Log Kow VALUES:	2.7 (measured), 2.8 (est.)	Mabey <i>et al.</i> (1982)
FINAL CHOSEN Log Kow VALUE:	2.7	

BASELINE UNCERTAINTY FACTOR FOR GUIDELINE DEVELOPMENT

IF Log Kow < 4.00, USE 1000

IF Log Kow ≥ 4.00, USE 10000

BASELINE UNCERTAINTY FACTOR:

1000

TABLE 2: ACUTE AQUATIC TOXICITY DATA FOR TOLUENE

SPECIES	LIFE STAGE	RESPONSE KEY (1)	pH	TEMP. (°C)	TEST CONDITIONS					EFFECT CONC. (mg/L)	DATA CODES KEY (2)	DATA QUALITY KEY (3)	REFERENCES
					D.O. (mg/L)	ALK. (mg/L)	HARD. (mg/L)						
VERTEBRATES Fathead minnow (<i>Pimephales promelas</i>)	30 days old	96h-LC50	8.3	25			80		30	F/M	PA	Devlin et al., 1982	
		96h-LC50	8.3	25			80		31	F/M	PA	Devlin et al., 1982	
		96h-LC50	8.3	25			80		28	F/M	PA	Devlin et al., 1982	
		96h-LC50	8.3	25			80		18	F/M	PA	Devlin et al., 1982	
	prolarvae	96h-LC50	8.3	25			80		36	F/M	PA	Devlin et al., 1982	
		96h-LC50	8.3	25			80		25	F/M	PA	Devlin et al., 1982	
		96h-LC50	8.3	25					27	F/M	PA	Devlin et al., 1982	
		96h-LC50	8.3	25			80		28	F/M	PA	Devlin et al., 1982	
	embryos	96h-LC50	8.3	25					72	F/M	PA	Devlin et al., 1982	
		96h-LC50	8.3	25			80		66	F/M	PA	Devlin et al., 1982	
		96h-LC50	8.3	25			80		59	F/M	PA	Devlin et al., 1982	
		96h-LC50	8.3	25			80		55	F/M	PA	Devlin et al., 1982	

KEY (1)	KEY (2)	KEY (3)
TLm = Median Tolerance Limit	S = Static	P = primary
LC = Lethal Concentration	F = Flowthrough	S = secondary
IC = Immobilization Concentration	U = Unmeasured Toxicant Concentration	T = tertiary
EC = Effects Concentration	M = Measured Toxicant Concentration	A = acute toxicity
	R = Renewed Static	C = chronic toxicity

TABLE 2: ACUTE AQUATIC TOXICITY DATA FOR TOLUENE

TEST CONDITIONS												
SPECIES	LIFE STAGE	RESPONSE KEY (1)	pH	TEMP. (°C)	D.O. (mg/L)	ALK. (mg/L)	HARD. (mg/L)	EFFECT CONC. (mg/L)	DATA CODES KEY (2)	DATA QUALITY KEY (3)	REFERENCES	
Fathead minnow (<i>Pimephales promelas</i>)	10-15 days old fry	96h-LC50	7.6-8.3	22±1				98-125	56.4	S/M	SA	Mayes et al., 1983
	30-35 days juvenile	96h-LC50	7.6-8.3	22±1				98-125	77.4	S/M	SA	Mayes et al., 1983
	60-100 days subadult	96h-LC50	7.6-8.3	22±1				98-125	54.0	S/M	SA	Mayes et al., 1983
	3.8-6.4 cm (1-2g)	96h-TLm	7.5	25	7.8	18	20	34.27	S/U	SA	Pickering & Henderson, 1986	
		96h-TLm	7.5	25	7.8	18	20	44	S/U	SA	Pickering & Henderson, 1986	
		96h-TLm	8.2	25	7.8	300	360	42.33	S/U	SA	Pickering & Henderson, 1986	
		96h-TLm	8.2	25	7.8	300	360	45	S/U	SA	Pickering & Henderson, 1986	
	adult	96h-LC50	7.7±0.35	20		38±20	33.8±19		12.6	S/U	SA	Liu et al., 1983
	31 days old	96h-LC50	7.89±0.03	24.7±0.2	6.9±0.44	43.4±1.25	45.4±0.8		36.2	F/M	SA	Geiger et al., 1986
KEY (1)												
TLm = Median Tolerance Limit												
LC = Lethal Concentration												
IC = Immobilization Concentration												
EC = Effects Concentration												
KEY (2)												
S = Static												
F = Flowthrough												
U = Unmeasured Toxicant Concentration												
M = Measured Toxicant Concentration												
R = Renewed Static												
KEY (3)												
P = primary												
S = secondary												
T = tertiary												
A = acute toxicity												
C = chronic toxicity												

TABLE 2: ACUTE AQUATIC TOXICITY DATA FOR TOLUENE

TEST CONDITIONS

SPECIES	LIFE STAGE	RESPONSE KEY (1)	pH	TEMP. (°C)	D.O. (mg/L)	ALK. (mg/L)	HARD. (mg/L)	EFFECT CONC. (mg/L)	DATA CODES KEY (2)	DATA QUALITY KEY (3)	REFERENCES
Fathead minnow (<i>Pimephales promelas</i>)	3.8-4.4 cm (1-2g)	48h-TLm	7.5	25	7.8	18	20	48.31	S/U	SA	Pickering & Henderson, 1986
		48h-TLm	8.2	25	7.8	300	360	58	S/U	SA	Pickering & Henderson, 1986
		24h-TLm	7.5	25	7.8	18	20	48.31	S/U	SA	Pickering & Henderson, 1986
		24h-TLm	8.2	25	7.8	300	360	58	S/U	SA	Pickering & Henderson, 1986
	30-31 days old	96h-LC50	7.8±0.02	25.5±0.35	8.4±0.4	48.7±0.17	39.7±0.9	31.7	F/M	PA	Geiger et al., 1990
Goldfish (<i>Carassius auratus</i>)	1-1.5 year old	96h-LC50	7.0±0.3	17-19	>7.0	71	80	22.8	F/M	PA	Brenniman et al., 1978
	(13-20 cm)	72h-LC50	7.0±0.3	17-19	>7.0	71	80	25.33	F/M	PA	Brenniman et al., 1978
	(20-80g)	48h-LC50	7.0±0.3	17-19	>7.0	71	80	27.82	F/M	PA	Brenniman et al., 1978
		24h-LC50	7.0±0.3	17-19	>7.0	71	80	41.59	F/M	PA	Brenniman et al., 1978
	3.8-4.4 cm (1-2g)	96h-TLm	7.5	25	7.8	18	20	57.88	S/U	SA	Pickering & Henderson, 1986

KEY (1)

TLm = Median Tolerance Limit
 LC = Lethal Concentration
 IC = Immobilization Concentration
 EC = Effects Concentration

KEY (2)

S = Static
 F = Flowthrough
 U = Unmeasured Toxicant Concentration
 M = Measured Toxicant Concentration
 R = Renewed Static

KEY (3)

P = primary
 S = secondary
 T = tertiary
 A = acute toxicity
 C = chronic toxicity

TABLE 2: ACUTE AQUATIC TOXICITY DATA FOR TOLUENE

SPECIES	LIFE STAGE	RESPONSE KEY (1)	pH	TEMP. (°C)	TEST CONDITIONS					EFFECT CONC. (mg/L)	DATA CODES KEY (2)	DATA QUALITY KEY (3)	REFERENCES
					D.O. (mg/L)	ALK. (mg/L)	HARD. (mg/L)	CONC. (mg/L)					
Goldfish (<i>Carassius auratus</i>)		96h-TLm	7.5	25	7.8	18	20	62	S/U	SA	Pickering & Henderson, 1966		
		48h-TLm	7.5	25	7.8	18	20	57.68	S/U	SA	Pickering & Henderson, 1968		
		24h-TLm	7.6	25	7.8	18	20	57.68	S/U	SA	Pickering & Henderson, 1968		
	6.2±0.7 cm (3.3±1.0g)	24h-TLm	7.8	20±1				58	S/M	SA	Bridle et al., 1979		
Coho salmon (<i>Oncorhynchus kisutch</i>)	Fry 0.3g	96h-LC50		7.8-10.4	11.0			5.48	F/M	PA	Moles et al., 1981		
	Fry 0.3g	96h-LC50						8.11	F/M	SA	Moles, 1980		
Rainbow trout (<i>Oncorhynchus mykiss</i>)	2.4g	96h-LC50	7.2-7.5	12±1		30-35	40-50	24	S/U	SA	Johnson & Finley, 1980		
		96h-LC50		12±1				5.8	R/M	PA	Galassi et al., 1988		

KEY (1)		KEY (2)		KEY (3)	
TLm	= Median Tolerance Limit	S	= Static	P	= primary
LC	= Lethal Concentration	F	= Flowthrough	S	= secondary
IC	= Immobilization Concentration	U	= Unmeasured Toxicant Concentration	T	= tertiary
EC	= Effects Concentration	M	= Measured Toxicant Concentration	A	= acute toxicity
		R	= Renewed Static	C	= chronic toxicity

TABLE 2: ACUTE AQUATIC TOXICITY DATA FOR TOLUENE

SPECIES	TEST CONDITIONS							EFFECT CONC. (mg/L)	DATA CODES KEY (2)	DATA QUALITY KEY (3)	REFERENCES
	LIFE STAGE	RESPONSE KEY (1)	pH	TEMP. (°C)	D.O. (mg/L)	ALK. (mg/L)	HARD. (mg/L)				
Bluegill sunfish (<i>Lepomis macrochirus</i>)	0.32g-1.2g	96h-LC50	6.7-7.8	22±1	7.0-8.8	28-34	32-48	13	S/M	SA	Buccafusco et al., 1981
	0.1g	96h-LC50	7.2-7.5	17±1	30-35	40-50	40-50	170	S/U	SA	Johnson & Finley, 1980
	1-2g	96h-TLm	7.5	25	7.8	18	20	24	S/U	SA	Pickering & Henderson, 1986
	(3.8-6.4 cm)	48h-TLm	7.5	25	7.8	18	20	24	S/U	SA	Pickering & Henderson, 1986
		24h-TLm	7.5	25	7.8	18	20	24	S/U	SA	Pickering & Henderson, 1986
Guppy (<i>Labiasia reticulatus</i>)	0.32-1.2g	24h-LO50	6.7-7.8	22±1	7.0-8.8	28-34	32-48	17	S/M	SA	Buccafusco et al., 1981
	1.9-2.5 cm	96h-TLm	7.5	25	7.8	18	20	59.3	S/U	SA	Pickering & Henderson, 1986
	(0.1-0.2g)	96h-TLm	7.5	25	7.8	18	20	86	S/U	SA	Pickering & Henderson, 1986
		48h-TLm	7.5	25	7.8	18	20	80.95	S/U	SA	Pickering & Henderson, 1986

KEY (1)	KEY (2)	KEY (3)
TLm = Median Tolerance Limit	S = Static	P = primary
LC = Lethal Concentration	F = Flowthrough	S = secondary
IC = Immobilization Concentration	U = Unmeasured Toxicant Concentration	T = tertiary
EC = Effects Concentration	M = Measured Toxicant Concentration	A = acute toxicity
	R = Renewed Static	C = chronic toxicity

TABLE 2: ACUTE AQUATIC TOXICITY DATA FOR TOLUENE

SPECIES	LIFE STAGE	RESPONSE KEY (1)	TEST CONDITIONS					EFFECT CONC. (mg/L)	DATA CODES KEY (2)	DATA QUALITY KEY (3)	REFERENCES
			pH	TEMP. (°C)	D.O. (mg/L)	ALK. (mg/L)	HARD. (mg/L)				
Guppy (<i>Lebistes reticulatus</i>)		24h-TLm	7.5	25	7.8	18	20	62.81	S/U	SA	Pickering & Henderson, 1988
		96h-LC50		21±1				28.2	R/M	PA	Gelassi et al., 1988
Channel catfish (<i>Ictalurus punctatus</i>)	0.1g	96h-LC50	7.2-7.5	22±1		30-35	40-50	240	S/U	SA	Johnson & Finley, 1980
Moaquillo fish (<i>Gambusia affinis</i>)		96h-TLm	7.5-8.6	17-22				1180	S/U	SA	Wallen et al., 1957
		24h-TLm	7.5-8.5	17-22				1340	S/U	SA	Wallen et al., 1957
		48h-TLm	7.5-8.5	17-22				1280	S/U	SA	Wallen et al., 1957
Golden orla (<i>Leuciscus idus</i>)		48h-LC50						70	S/U	SA	Juhnke & Lüdemann, 1978
		48h-LC50						422	S/U	SA	Juhnke & Lüdemann, 1978

KEY (1)	KEY (2)	KEY (3)
TLm = Median Tolerance Limit	S = Static	P = primary
LC = Lethal Concentration	F = Flowthrough	S = secondary
IC = Immobilization Concentration	U = Unmeasured Toxicant Concentration	T = tertiary
EC = Effects Concentration	M = Measured Toxicant Concentration	A = acute toxicity
	R = Renewed Static	C = chronic toxicity

TABLE 2: ACUTE AQUATIC TOXICITY DATA FOR TOLUENE

SPECIES	LIFE STAGE	RESPONSE KEY (1)	pH	TEMP. (°C)	TEST CONDITIONS					EFFECT CONC. (mg/L)	DATA CODES KEY (2)	DATA QUALITY KEY (3)	REFERENCES
					D.O. (mg/L)	ALK. (mg/L)	HARD. (mg/L)						
INVERTEBRATES													
Cladoceran (<i>Daphnia magna</i>)	<24h old	48h-EC50	6.7-8.1	22±1	6.5-9.1			72±6	310	S/U	SA		LeBlanc, 1980
		48h-EC60	7.7±0.35	20		38±20		33.8±19	20.2	S/M	SA		Liu et al., 1983
	<24h old	24h-LC50	6.7-8.1	22±1	6.5-9.1			72±6	310	S/U	SA		LeBlanc, 1980
		No discernable effect	6.7-8.1	22±1	6.5-9.1			72±6	28	S/U	SA		LeBlanc, 1980
	<2d old	48h-EC50(immob.)		22±1					14.93		S/M		
	24h old	24h-EC50(immob.)							7	S/M	PA		Gafeel et al., 1988
	<72h old	48h-EC50(immob.)	6.0-7.0	23±2	5.0-9.0				11.5	S/U	SA		Bobra et al., 1983

KEY (1)		KEY (2)		KEY (3)	
TLm	Median Tolerance Limit	S	Static	P	primary
LC	Lethal Concentration	F	Flowthrough	S	secondary
IC	Immobilization Concentration	U	Unmeasured Toxicant Concentration	T	tertiary
EC	Effects Concentration	M	Measured Toxicant Concentration	A	acute toxicity
		R	Renewed Static	C	chronic toxicity

TABLE 3: CHRONIC AQUATIC TOXICITY DATA FOR TOLUENE

TEST CONDITIONS

SPECIES	LIFE STAGE	RESPONSE KEY (1)	pH	TEMP. (°C)	D.O. (mg/L)	ALK. (mg/L)	HARD. (mg/L)	EFFECT CONC. (mg/L)	DATA CODES KEY (2)	DATA QUALITY KEY (3)	REFERENCES
VERTEBRATES											
Rainbow trout (<i>Oncorhynchus mykiss</i>)	Embryo exposed from fertilization to: hatch	23d-LC50	7.8±0.02	14.3±0.2	9.8±0.1		108.3±1.2	0.03	F/M	PC	Black et al., 1982
	4d posthatch	27d-LC50	7.8±0.02	14.3±0.2	9.8±0.1		108.3±1.2	0.02	F/M	PC	Black et al., 1982
Coho Salmon (<i>Oncorhynchus kisutch</i>)	Fry	40d—sign. decr. wet & dry wt.		7.8-10.4				2.78	F/M	PC	Moles et al., 1981
		40d—sign. decr. wet & dry wt. & length		7.8-10.4				5.00	F/M	PC	Moles et al., 1981
Goldfish (<i>Carassius auratus</i>)	1-1.5 year old (13-20 cm 20-80 g)	30d-LC50	7.0±0.3	17-19	>7.0	71	80	14.58	F/M	PC	Brenneman et al., 1978
Guppy (<i>Poecilia reticulata</i>)	2-3 months old	14d-LC50		22±1	6		25	68.3	R/U	SC	Kühmann, 1981
<div> <div>KEY (1)</div> <div> TLm = Median Tolerance Limit LC = Lethal Concentration IC = Immobilization Concentration EC = Effects Concentration </div> </div> <div> <div>KEY (2)</div> <div> S = Static F = Flowthrough U = Unmeasured Toxicant Concentration M = Measured Toxicant Concentration R = Renewed Static </div> </div> <div> <div>KEY (3)</div> <div> P = primary S = secondary T = tertiary A = acute toxicity C = chronic toxicity </div> </div>											

TABLE 3: CHRONIC AQUATIC TOXICITY DATA FOR TOLUENE

TEST CONDITIONS

SPECIES	LIFE STAGE	RESPONSE KEY (1)	pH	TEMP. (°C)	D.O. (mg/L)	ALK. (mg/L)	HARD. (mg/L)	EFFECT CONC. (mg/L)	DATA CODES KEY (2)	DATA QUALITY KEY (3)	REFERENCES
Fathead minnow (<i>Pimephales promelas</i>)	30-35d old	8d-LC50		25±2	6.3-8.2	38-44	43-48	44.1	F/M	PC	Hall et al., 1984, 1989
Madaka (<i>Oryzias latipes</i>)	embryos	96h-LC50		25				54	S/U	SC	Stoss & Haines, 1979
Salamander (<i>Ambystoma gracile</i>)	Embryos exposed from fertil- ization to: hatch 4d posthatch	5.6d-LC50 9.6d-LC50	7.7±0.02 7.7±0.02	20.2±0.5 20.2±0.5	7.5±0.1 7.5±0.1		98.9±0.9 98.9±0.9	1.09 0.65	F/M F/M	PC PC	Black et al., 1982 Black et al., 1982
Leopard frog (<i>Rana pipiens</i>)	Embryos exposed from fertil- ization to: hatch 4d posthatch	5d-LC50 9d-LC50	7.7±0.02 7.7±0.02	20.2±0.5 20.2±0.5	7.5±0.1 7.5±0.1		98.9±0.9 98.9±0.9	0.51 0.39	F/M F/M	PC PC	Black et al., 1982 Black et al., 1982

KEY (1) = Median Tolerance Limit
 TLm = Lethal Concentration
 LC = Immobilization Concentration
 EC = Effects Concentration

KEY (2)
 TLm = Stallo
 F = Flowthrough
 U = Unmeasured Toxicant Concentration
 M = Measured Toxicant Concentration
 R = Renewed Stallo

KEY (3)
 P = primary
 S = secondary
 T = tertiary
 A = acute toxicity
 C = chronic toxicity

TABLE 3: CHRONIC AQUATIC TOXICITY DATA FOR TOLUENE

TEST CONDITIONS

SPECIES	LIFE STAGE	RESPONSE KEY (1)	pH	TEMP. (°C)	D.O. (mg/L)	ALK. (mg/L)	HARD. (mg/L)	EFFECT CONC. (mg/L)	DATA CODES KEY (2)	DATA QUALITY KEY (3)	REFERENCES
INVERTEBRATES											
Cladoceran (<i>Daphnia magna</i>)	<24h old	16d-LC50 (QSAR est.)						3.75		SC	Hermans et al., 1984
OTHER ORGANISMS											
Rotatoria (<i>Dicranophorus toxicipatus</i>)		6d-EC40 (cell culture growth)						173	S/U	SC	Erben, 1978
ALGAE											
<i>Chlamydomonas angulosa</i>		3h-EC50 (photosynth.)						134		SC	Hutchinson et al., 1980
<i>Chlorella vulgaris</i>		3h-EC50 10d-EC50 (growth)						207 245		SC SC	Hutchinson et al., 1980 Kause & Hutchinson, 1975
<i>Scenedesmus quadricauda</i>		cell multiplication inhibition test toxicity threshold						>400		TC	Brüggemann & Kuhn, 1980
<i>Solenastrium capricornutum</i>		72h-EC50 (growth)						12.5	S/M	SC	Galassi et al., 1988
<div> <div>KEY (1)</div> <div> T_{lm} = Median Tolerance Limit LC = Lethal Concentration IC = Immobilization Concentration EC = Effects Concentration </div> </div> <div> <div>KEY (2)</div> <div> S = Stallo F = Flowthrough U = Unmeasured Toxicant Concentration M = Measured Toxicant Concentration R = Renewed Static </div> </div> <div> <div>KEY (3)</div> <div> P = primary S = secondary T = tertiary A = acute toxicity C = chronic toxicity </div> </div>											

TABLE 3: CHRONIC AQUATIC TOXICITY DATA FOR TOLUENE

TEST CONDITIONS

SPECIES	LIFE STAGE	RESPONSE KEY (1)	pH	TEMP. (°C)	D.O. (mg/L)	ALK. (mg/L)	HARD. (mg/L)	EFFECT CONC. (mg/L)	DATA CODES KEY (2)	DATA QUALITY KEY (3)	REFERENCES
BACTERIA											
<i>Pseudomonas putida</i>		cell multiplication inhibition test toxicity threshold						29		TC	Bringmann & Kuhn, 1980
PROTOZOA											
<i>Tetrahymena pyriformis</i>		24h-LC100						550	S/U	TC	Schultz et al., 1978
<i>Enosiphon sulcatum</i>		cell multiplication inhibition test toxicity threshold						>450		TC	Bringmann & Kuhn, 1980
KEY (1)	KEY (2)	KEY (3)									
TLm = Median Tolerance Limit	S = Static	P = primary									
LC = Lethal Concentration	F = Flowthrough	S = secondary									
IC = Immobilization Concentration	U = Unmeasured Toxicant Concentration	T = tertiary									
EC = Effects Concentration	M = Measured Toxicant Concentration	A = acute toxicity									
	R = Renewed Static	C = chronic toxicity									

Table 4: UNCERTAINTY FACTOR WORKSHEET

CHEMICAL:	CAS No.	CONCENTRATION UNITS
TOLUENE	108-88-3	mg/L

Test Conditions		Species (life stage)	Toxicity End Point	Effect conc.	Data Codes ¹	Data Type ²	Calibration Factor	Reference
ACUTE	VERTEBRATE	Fathead minnow, 30d	96h-LC50	18	F/M	1°	0.8	Devlin <i>et al.</i> 1982
		Rainbow trout	96h-LC50	5.8	R/M	1°	0.8	Galassi <i>et al.</i> 1988
		Coho salmon, fry	96h-LC50	5.46	F/M	1°	0.8	Moles <i>et al.</i> 1981
	INVERT.	<i>Daphnia magna</i>	24h-EC50 (imm.)	7.0	R/M	1°	0.8	Galassi <i>et al.</i> 1988

CHRONIC	VERTEBRATE	Goldfish, 1.0-1.5y old	30d-LC50	14.58	F/M	1°	0.5	Brenniman <i>et al.</i> 1976
		Rainbow trout, embryo	27d-LC50	0.02	F/M	1°	0.5	Black <i>et al.</i> 1982
		Coho salmon, fry	40d-sign. gr. red	2.76	F/M	1°	0.5	Moles <i>et al.</i> 1981
	INVERT.	<i>Daphnia magna</i> , <24h	16d-LC50 (OSAR)	3.75		3°	0.8	Hermens <i>et al.</i> 1984
		<i>Dicranophorus farcipatus</i>	6d-EC40 (gr)	173	S/U	2°	0.7	Erben 1978
	PLANT	<i>Selenastrum capricornutum</i>	72h-EC50 (gr.)	12.5	R/M	2°	0.9	Galassi <i>et al.</i> 1988

CALCULATION OF FINAL UNCERTAINTY FACTOR:

Since Log Kow < 4.00, The Baseline Uncertainty Factor = 1000

Baseline Uncertainty Factor X Calibration Factors (maximum number = 11)

$$1000 \times 0.8 \times 0.8 \times 0.8 \times 0.8 \times 0.5 \times 0.5 \times 0.5 \times 0.8 \times 0.7 \times 0.9 \times \boxed{} = \boxed{26} \text{ FINAL UNCERTAINTY FACTOR}$$

CRITICAL VALUE ÷ FINAL UNCERTAINTY FACTOR = PWQG

$$= \frac{0.02}{26} = 0.0008 \text{ mg/L}$$

¹ Assign 2 DATA CODES, one from each of the following rows:
 S = static R = static/renewal F = flowthrough
 U = unmeasured nominal conc. M = measured conc.

² DATA TYPE:
 1° = Primary 2° = Secondary 3° = Simulated Data

FIG. 1: TOXICITY SUMMARY AND GUIDELINE DERIVATION GRAPH

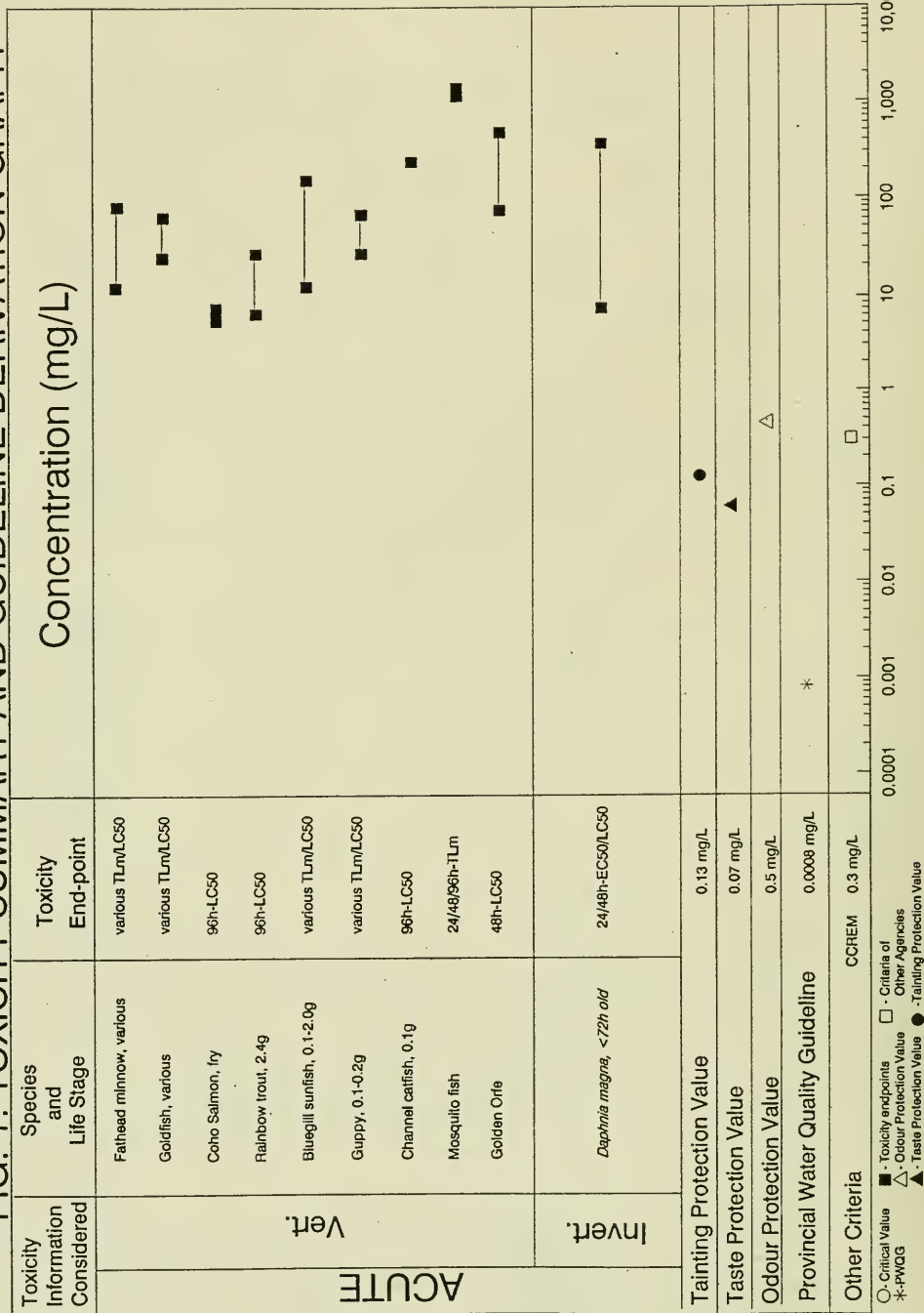


FIG. 1: TOXICITY SUMMARY AND GUIDELINE DERIVATION GRAPH

